

# Oak or chestnut tannin dose responses on silage pH, proteolysis and *in vitro* digestibility in laboratory-scale silos

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**Description of the subject.** This short note documents the use of hydrolyzable tannins as silage additives to reduce proteolysis thanks to a laboratory-scale ensiling method.

**Objectives.** To study oak (OTE) and chestnut tannin extract (CTE) dose responses on chemical composition, pH and ammoniacal nitrogen (N-NH<sub>3</sub>) content of silage.

**Method.** A mixture of cocksfoot, white and red clovers was ensiled in vacuum packs, with OTE or CTE at doses of 0, 10, 30, 50 and 70 g·kg<sup>-1</sup> DM.

**Results.** Hydrolyzable tannin extracts decreased N-NH<sub>3</sub> content of silage up to 18% ( $p < 0.05$ ). For the investigated range of doses, OTE induced a linear decrease of N-NH<sub>3</sub> content ( $R^2 = 0.76$ ) whereas CTE resulted in a quadratic decrease ( $R^2 = 0.68$ ). High doses of tannin extracts reduced *in vitro* organic matter digestibility (OMD) by 3% ( $p < 0.05$ ).

**Conclusions.** Both tannins reduced proteolysis in silos but highest doses induced a decrease in OMD.

**Keywords.** Additives, ammonia, fermentation.

## Effets des doses des tanins de chêne ou de châtaignier sur le pH, la protéolyse et la digestibilité *in vitro* d'ensilages en micro-silos

**Description du sujet.** Cette note de recherche documente, en micro-silos, l'utilisation de tannins hydrolysables comme additifs d'ensilage pour réduire la protéolyse.

**Objectifs.** Étudier les effets dose d'extraits de tannin de chêne (OTE) et de châtaignier (CTE) sur la composition chimique et la conservation d'ensilage d'herbe.

**Méthode.** Un mélange de dactyle, trèfles blanc et violet a été ensilé en sachets sous vide, avec des doses de 0, 10, 30, 50 et 70 g·kg<sup>-1</sup> MS d'OTE et CTE.

**Résultats.** Les tannins hydrolysables ont réduit la concentration en azote ammoniacal (N-NH<sub>3</sub>) des ensilages jusqu'à 18 % ( $p < 0,05$ ). Pour les doses étudiées, l'OTE a montré une diminution linéaire du N-NH<sub>3</sub> ( $R^2 = 0,76$ ), tandis que le CTE a provoqué une réduction quadratique ( $R^2 = 0,68$ ). Les fortes doses de tannins ont réduit la digestibilité *in vitro* de la matière organique (DMO) de 3 % ( $p < 0,05$ ).

**Conclusions.** Les deux tannins réduisent la protéolyse dans l'ensilage, mais de fortes doses diminuent la DMO.

**Mots-clés.** Additif, ammoniac, fermentation.

## 1. INTRODUCTION

Forages are a main source of proteins for ruminants, especially legume forages. Rich in degradable proteins, these forages are highly susceptible to proteolysis during ensiling (Albrecht & Beauchemin, 2003).

However, proteolysis intensity is negatively correlated to tannin concentration in silage (Albrecht & Muck, 1991). These natural polyphenols present in some plants make complexes with proteins and can protect them against degradation in silo but also in rumen (Piluzza et al., 2014). Several studies showed a reduction of

non-protein N and/or ammonia in silo when adding hydrolyzable tannin as silage additive (Cavallarin et al., 2002; Tabacco et al., 2006; Herremans et al., 2018). In order to study silages, cheap and convenient models of real-size silos are needed. Vacuum packs are commonly used because of their interesting cost and flexible utilization. Comparisons with glass tubes have shown marginal or no differences regarding fermentation parameters for grass and legume silages (Johnson et al., 2005). The objective of this work was to study the effect of increasing doses of oak (OTE) or chestnut tannin extracts (CTE) on conservation of grass/legume mixed silage in laboratory-scale vacuum packs.

## 2. MATERIALS AND METHODS

A mature stand of cocksfoot (*Dactylis glomerata*), white clover (*Trifolium repens*) and red clover (*Trifolium pratense*) was harvested in Gembloux, Belgium, as a second cut in July 2018. Forage contained 615 g of cocksfoot and 385 g of mixed white and red clovers per kg of DM. Forage was cut and chopped before being ensiled in plastic packs (10 l capacity, Brouwland, Beverlo, Belgium) according to two additive treatments: OTE or CTE. For each tannin type, five doses were investigated (0, 10, 30, 50 and 70 g·kg<sup>-1</sup> forage DM). OTE and CTE extracts contained 468 ± 6 g·kg<sup>-1</sup> DM and 492 ± 7 g·kg<sup>-1</sup> DM respectively of total tannins (dosed by methyl cellulose precipitable method). Each treatment-dose combination was replicated five times and packs were filled with 600 g of fresh forage. Where appropriate, powdered tannin

extracts were added and mixed directly in packs. The 50 packs were sealed with a vacuum packaging machine (model UNICA GAS, Lavezzini, Fiorenzuola, Italy) and stored at room temperature. After 28 days, laboratory-scale silos were opened and homogenized. Forages were oven-dried at 60 °C until constant weight and ground in a Cyclotec mill (1 mm screen, FOSS, Hillerød, Denmark). Chemical composition and *in vitro* organic matter digestibility (OMD; de Boever et al., 1986) were determined by near infrared reflectance spectroscopy (database of Minet et al., 2016; XDS-system spectrometer, FOSS, Hillerød, Denmark). Measurement of pH was carried out with a conventional glass electrode in the water extract. Silage N-NH<sub>3</sub> content was measured in the acidified (pH < 3) water extract by the Kjeldahl method (Kjeltec, FOSS, Hillerød, Denmark). Using Minitab® Statistical Software, all data were processed through a general linear model with tannin type and dose as fixed factors. Dunnett post-hoc test was used to compare dose means to dose 0 mean. A regression analysis on tannin dose response of N-NH<sub>3</sub> content was performed with Minitab Assistant. The regression model with the highest-order significant term (*p* < 0.05) was considered as the best.

## 3. RESULTS

After ensiling, DM content reached 314 g·kg<sup>-1</sup> on average. Tannin dose significantly affected all composition parameters (**Table 1**, *p* < 0.05). Dry matter (DM) and crude protein (CP) content showed significant but minor variations. From a dose of 30 g·kg<sup>-1</sup> DM, acid detergent fibre (ADF) fell when increasing tannin

**Table 1.** Chemical composition, digestibility and conservation parameters of silages according to tannin type and dose — *Composition chimique, digestibilité et paramètres de conservation des ensilages en fonction du type de tannin et de la dose.*

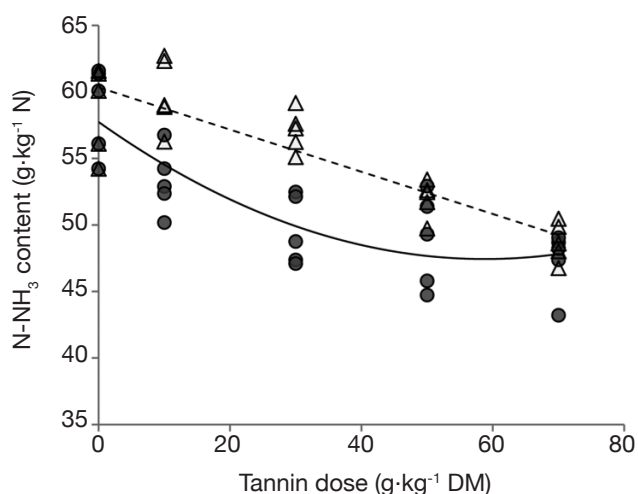
Parameter	Tannin type (t)		Dose (d)					SEM	p-value		
	Chestnut	Oak	0	10	30	50	70		t	d	t x d
DM (g·kg <sup>-1</sup> )	314.1	315.2	314.6	313.0	314.4	312.6	318.7*	0.53	0.205	<0.001	0.132
CP (g·kg <sup>-1</sup> DM)	173.1	174.1	170.9	172.1	175.5*	174.1*	175.4*	0.40	0.157	<0.001	0.385
ADF (g·kg <sup>-1</sup> DM)	299.3	296.1	302.6	301.5	296.7*	295.2*	292.5*	0.71	<0.001	<0.001	0.087
ADL (g·kg <sup>-1</sup> DM)	28.7	28.5	23.8	26.4*	28.4*	32.5*	32.9*	0.54	0.759	<0.001	0.600
OMD (%)	81.8	82.4	83.0	82.7	82.5	81.2*	81.3*	0.17	0.044	<0.001	0.341
pH	5.3	5.3	5.3	5.2	5.3	5.2	5.3	0.01	0.572	0.056	0.746
N-NH <sub>3</sub> (g·kg <sup>-1</sup> N)	51.5	55.3	58.7	56.6	53.3*	50.4*	48.0*	0.72	<0.001	<0.001	0.100

DM: dry matter — *matière sèche*; CP: crude protein — *protéines brutes*; ADF: acid detergent fibre — *fibres au détergent acide*; ADL: acid detergent lignin — *lignine au détergent acide*; OMD: *in vitro* organic matter digestibility — *digestibilité in vitro de la matière organique*; SEM: standard error of the mean — *erreur standard de la moyenne*; In the same line, values marked with an asterisk (\*) are significantly different from the control (Dunnett contrast test, compared to dose 0, *p* < 0.05) — *Sur une même ligne, les valeurs marquées d'une astérisque (\*) sont significativement différentes du témoin (test de Dunnett, comparaison à la dose 0, p < 0,05).*

dose whereas acid detergent lignin (ADL) increased. ADF was the only composition parameter influenced by the type of tannin, showing a lower level with OTE compared to CTE. OMD was affected by tannin from a dose of 50 g·kg<sup>-1</sup> forage DM. CTE induced a lower OMD than OTE ( $p < 0.05$ ). Tannins did not influence pH but ammoniacal nitrogen content was lower in CTE than in OTE silages (-7%). At 30 g·kg<sup>-1</sup> forage DM, tannin extracts reduced N-NH<sub>3</sub> content by 9% compared to 14% and 18% reductions at 50 and 70 g·kg<sup>-1</sup> forage DM respectively. No interactions were found between tannin type and dose. N-NH<sub>3</sub> content was negatively correlated to tannin dose (**Figure 1**). N-NH<sub>3</sub> content decreased linearly with OTE dose ( $R^2 = 0.76$ ) whereas it seems to decrease along a quadratic relation with CTE ( $R^2 = 0.68$ ). With CTE, the linear model showed a lower  $R^2$  ( $y = 56.14 - 0.1440 x$ ;  $p < 0.001$ ;  $R^2 = 0.58$ ).

#### 4. DISCUSSION

As tannin contents higher than 50 g·kg<sup>-1</sup> forage DM are known to reduce intake and protein digestibility (Piluzza et al., 2014), our highest dose was used for scientific interest only. According to Hagerman et al. (1992), hydrolyzable tannins such as oak or chestnut tannins should not affect feed digestibility because of



**Figure 1.** N-NH<sub>3</sub> content of silages (g·kg<sup>-1</sup> N, mean of five replications) in relation to tannin dose — *Concentration en N-NH<sub>3</sub> des ensilages (g·kg<sup>-1</sup> N, moyenne de cinq répétitions) en fonction de la dose de tannin.*

△: Oak tannin, dotted regression line,  $y = 60.34 - 0.1586 x$ ;  $p < 0.001$ ;  $R^2 = 0.76$  — *Tannin de chêne, ligne de régression pointillée,  $y = 60,34 - 0,1586 x$ ;  $p < 0,001$ ;  $R^2 = 0,76$* ; ●: Chestnut tannin, plain regression line,  $y = 57.74 - 0.3506 x + 0.002982 x^2$ ;  $p < 0.001$ ;  $R^2 = 0.68$  — *Tannin de châtaignier, ligne de régression continue,  $y = 57,74 - 0,3506 x + 0,002982 x^2$ ;  $p < 0,001$ ;  $R^2 = 0,68$ .*

their quick degradation in the gut. It seems more likely that the rise observed in ADL content would reflect the presence of lignin in the commercial tannin extracts, as they are wood by-products. This presence could explain most of the OMD decrease.

The N-NH<sub>3</sub> reduction supports the hypothesis that tannin-protein complexes are stable at pH 5 and that tannins reduce proteolysis in this environment. The quadratic relation observed with CTE seems to be confirmed by Tabacco et al. (2006), showing a higher decrease of N-NH<sub>3</sub> content with 40 g·kg<sup>-1</sup> relative to 20 g·kg<sup>-1</sup> forage DM but no differences between 40 and 60 g·kg<sup>-1</sup> forage DM doses of CTE in silages. Tabacco et al. (2006) explained the absence of further significant N-NH<sub>3</sub> reduction by an already complete inhibition of some silage microorganisms at 40 g·kg<sup>-1</sup> DM. The linear relation observed with OTE could be due to a threshold dose higher than the investigated doses. However, the hypothesis of a quadratic relation between hydrolysable tannin dose and silage N-NH<sub>3</sub> content should be confirmed by an experiment with higher tannin doses because of the short plateau observed with CTE and its absence with OTE at the tested doses. Globally, CTE had a slightly higher beneficial effect on silage proteolysis than OTE. This is probably due to the specificity of chemical bonds between tannins and proteins, which can differ according to tannin and protein structures. This specificity also induces an interaction between tannin and forage composition, as already shown in a previous work (Herremans et al., 2018).

#### 5. CONCLUSIONS

In order to improve protein conservation in silage, the best dose investigated in the present study seems to be around 30 g·kg<sup>-1</sup> forage DM of hydrolyzable tannin extract in silage. Lower dose prevents tannins from lowering proteolysis and doses from 50 g·kg<sup>-1</sup> DM seem to reduce OMD and probably dry matter intake. More research is needed on the dose range between 10 and 50 g·kg<sup>-1</sup> DM as well as on different forage compositions.

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