

# Effect of temperature on growth and fatty acids profile of the biodiesel producing microalga *Scenedesmus acutus*

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**Description of the subject.** The present study examined the effect of temperature (15, 20, 25, 30, 35 and 40 °C) on biomass, esterified fatty acids content and fatty acid productivity of *Scenedesmus acutus*.

**Objectives.** This work aimed to study the effect of variation in temperature on lipid productivity and fatty acid profiles of *S. acutus* as a feedstock for biodiesel production.

**Method.** The alga was grown under different temperatures and its biomass, as well as fatty acid content and composition, were determined.

**Results.** The maximum growth rate of *S. acutus* was achieved at 30 °C, but there was no significant difference in biomass productivity at 25 and 30 °C (0.41 and 0.42 g·l<sup>-1</sup>·d<sup>-1</sup>), respectively. The highest fatty acid content (104.1 mg·g<sup>-1</sup> CDW) was recorded at low temperature (15 °C) and decreased with increasing temperature. As a result of high biomass production, fatty acids productivity showed the highest values (41.27 and 42.10 mg·l<sup>-1</sup>·d<sup>-1</sup>) at 25 and 30 °C, respectively. The proportion of saturated and mono-unsaturated fatty acids increased from 13.72 to 23.79% and from 11.13 to 33.10% of total fatty acids when the incubation temperature was raised from 15 to 40 °C, respectively. The increase of temperature from 15 to 40 °C decreased the poly-unsaturated fatty acids from 75.15% to 43.10% of total fatty acids, respectively.

**Conclusions.** The present study concluded that incubation temperature was a critical parameter for quantitative and qualitative fatty acid compositions of *S. acutus*. In addition, the type and proportion of individual fatty acids, which interfere with biodiesel quality, can be modified using different incubation temperatures in order to meet the biodiesel international standards.

**Keywords.** Biomass, *Scenedesmus*, biodiesel, fatty acids, temperature, productivity.

## Effet de la température d'incubation sur la croissance et le profil en acides gras de la microalgue *Scenedesmus acutus* à mettre en rapport avec une production potentielle de biodiesel

**Description du sujet.** L'étude examine l'effet de la température d'incubation (15, 20, 25, 30, 35 et 40 °C) sur la biomasse, la productivité en acides gras de la microalgue *Scenedesmus acutus*, potentielle productrice de biodiesel.

**Objectifs.** Le travail a pour objectif d'étudier l'effet d'une variation de température sur le contenu lipidique de *S. acutus*, la productivité lipidique et la production de biomasse de l'algue.

**Méthode.** L'algue a été cultivée sous différentes températures de croissance et la teneur en lipides, la composition en acides gras et la biomasse de *S. acutus* ont été déterminés.

**Résultats.** L'incubation de *S. acutus* à 30 °C donne la meilleure croissance, bien qu'il n'y ait pas de différence significative pour la production de biomasse entre 25 et 30 °C (0,41 et 0,42 g·l<sup>-1</sup>·j<sup>-1</sup>, respectivement). La plus haute teneur en acides gras (104,1 mg·g<sup>-1</sup> CDW) a été obtenue à 15 °C et décroît lorsqu'on augmente la température. En raison de la forte production de biomasse, la productivité des acides gras a montré les valeurs les plus élevées à 25 et 30 °C (41,27 et 42,10 mg·l<sup>-1</sup>·j<sup>-1</sup>, respectivement). En ce qui concerne le profil en acides gras, les proportions d'acides gras saturés et mono-insaturés se sont accrues, tandis que celle des acides gras poly-insaturés a chuté avec l'augmentation de la température d'incubation. Les proportions d'acides gras saturés et mono-insaturés sont montées respectivement de 13,72 à 23,79 % et de 11,13 à 33,10 % dans la représentation des acides gras totaux, lorsque la température d'incubation est passée de 15 à 40 °C, tandis que pour ce même écart thermique, la proportion des acides gras poly-insaturés au sein des acides gras totaux a diminué de 75,15 % à 43,10 %.

**Conclusions.** On peut conclure de la présente étude que la température d'incubation est un paramètre critique pour la composition quantitative et qualitative des acides gras de *S. acutus*. De plus, le type et la proportion de chaque acide gras sont des paramètres qui interfèrent dans la qualité du biodiesel, mais qui peuvent être modifiés par la température d'incubation de manière à rencontrer les normes internationales du biodiesel.

**Mots-clés.** Biomasse, *Scenedesmus*, biodiésel, acide gras, température, productivité.

## 1. INTRODUCTION

Contemporary global energy usage is based on utilization of fossil fuels including natural gas, coal and oil. Barbir (2009) concluded that the use of fossil fuels has several problems, such as:

- pollution at local, regional and global scales;
- risk of complete depletion of fossil fuel energy, as the worldwide fossil oil reserves will be exhausted in shorter than 30 years due to the quick development of anthropogenic activities and overconsumption (Abomohra et al., 2016);
- increasing of greenhouse gas emissions (*i.e.* NO<sub>x</sub>, CO<sub>2</sub> and SO<sub>x</sub>) that cause global warming and climate problems.

Biofuel (fuel derived from biomass) receives considerable attention because it is a renewable, biodegradable and non-toxic fuel (Mutanda et al., 2010). Biodiesel from microalgae is a promising renewable energy that might completely replace the fossil diesel without influencing the human food supply (Chisti, 2008). In fact, algae have a much higher yield as a biodiesel feedstock than crop plants (Abomohra et al., 2014). The annual oil yield of algae is 7 to 31 times larger than palm oil, for the same given land area, due to their ability to accumulate lipids and their very high actual photosynthetic yield (Li et al., 2008; Abomohra et al., 2014). The total yield of biodiesel depends on the lipid content of the algal strain and also on its growth rate. For biodiesel production, the economic biomass production of microalgae has to be taken into consideration, so microalgal species with a high lipid content and a high cell growth are used (Lv et al., 2010). Abomohra et al. (2013) reported two categories of microalgae for high lipid production:

- high lipid content (43%) with low growth rate (30 mg·l<sup>-1</sup>·d<sup>-1</sup>), such as *Botryococcus braunii*,
- high growth rate (250 mg·l<sup>-1</sup>·d<sup>-1</sup>) with low lipid content (15%) such as *Scenedesmus obliquus*.

Several studies demonstrated that the quantity and quality of intracellular lipids and the cellular growth rate can vary as a result of changes in growth conditions including light intensity, temperature, CO<sub>2</sub> concentrations, nitrogen or phosphate limitation, silicon deficiency, and iron supplementation (Tzovenis et al., 2003; Liu et al., 2008; Griffiths & Harrison, 2009; Ho et al., 2010; El-Sheekh et al., 2013; Kumar

et al., 2014). Most of the previous studies investigated the impact of those factors on cell growth and/or lipid content of various microalgal species separately. However, to get better understanding of the relationship between cell growth and lipid accumulation, they have to be measured as biomass and lipid productivities. Furthermore, among the factors mentioned above, temperature is a sensitive limiting factor for microalgal growth and metabolic activities. Moreover, it is an easy-to-control factor in the practical operation of microalgae cultivation. In this study, the effects of different incubation temperatures on *S. acutus* were carried out in a batch cultivation mode. The variation in fatty acids content, fatty acids productivity, and fatty acids profiles in response to different temperatures was measured. The influence of incubation temperature on cell growth and biomass production was also assessed.

## 2. MATERIALS AND METHODS

### 2.1. Algal strain and cultivation conditions

*Scenedesmus acutus* was isolated from municipal wastewater at Tanta, Egypt, and cultivated axenically in 11 Erlenmeyer flasks with 700 ml KC medium (Kessler & Czygan, 1970) at an initial OD<sub>680</sub> of 0.05 nm. Sterile filtered air enriched with 3%<sup>680</sup> CO<sub>2</sub> (v/v) was continuously applied to the cultures. Cultures were illuminated from the top using tubular fluorescent lamps (PHILIPS Master TL-D 85 W / 840) with light intensity of 130 ± 10 μmol photons·m<sup>-2</sup>·s<sup>-1</sup> at the surface of the flasks with a photoperiod of 14:10 h light:dark at different incubation temperatures (15, 20, 25, 30, 35 and 40 °C). The optical density of the algal suspension was measured at 680 nm every other day. Dry weight and esterified fatty acids (EFAs) were measured after 22 days of incubation and allowed calculation of biomass and fatty acids productivities. Cell number showed a strong positive correlation with the optical density (R<sup>2</sup> = 0.991), which can be calculated from the equation:

$$\text{Cell number } (\times 10^6 \text{ cell}\cdot\text{ml}^{-1}) = 1.423 \times OD_{680} - 0.002 \quad (R^2 = 0.991)$$

### 2.2. Biomass assay

Algal growth was monitored by measuring the optical density at 680 nm (OD<sub>680</sub>) and by determination of the

cellular dry weight (CDW). From the values of CDW, biomass productivity was calculated according to Abomohra et al. (2013) as follows:

$$\text{Biomass productivity (g CDW} \cdot \text{l}^{-1} \cdot \text{d}^{-1}) = \text{CDW}_L - \text{CDW}_E \cdot (\text{t}_L - \text{t}_E)^{-1}$$

where  $\text{CDW}_E$  and  $\text{CDW}_L$  represent the CDW ( $\text{g} \cdot \text{l}^{-1}$ ) at the days of early exponential phase ( $\text{t}_E$ ) and late exponential phase ( $\text{t}_L$ ), respectively.

### 2.3. Lipid extraction

To analyze the cellular fatty acids composition, 5 ml aliquots of each culture were collected after 22 days of incubation at specified times. Lipids were extracted following the method of Bligh & Dyer (1959). Prior to extraction, trionadecanoylglycerol was added to the samples as an internal standard for esterified fatty acids.

### 2.4. Fatty acids profile

Esterified fatty acids (EFA) from the extract of intracellular lipids were transmethylated for GC analysis as previously described (Scharnewski et al., 2008; Kaczmarzyk & Fulda, 2010). The fatty acids methyl esters were subjected to GC/FID analysis. The GC analysis was performed with a Varian 3900 GC-system equipped with a capillary column (Select Fame, 50 m  $\times$  0.25 mm; Varian). Fatty acids content ( $\text{mg} \cdot \text{g}^{-1}$  CDW) was calculated; in addition, fatty acids productivity was calculated according to Abomohra et al. (2013):

$$\text{Fatty acids productivity (mg} \cdot \text{l}^{-1} \cdot \text{d}^{-1}) = (\text{FA}_L - \text{FA}_E) \cdot (\text{t}_L - \text{t}_E)^{-1}$$

where  $\text{FA}_E$  and  $\text{FA}_L$  represent the total fatty acids content ( $\text{mg} \cdot \text{l}^{-1}$ ) at the days of early exponential phase ( $\text{t}_E$ ) and late exponential phase ( $\text{t}_L$ ), respectively.

### 2.5. Statistical analysis

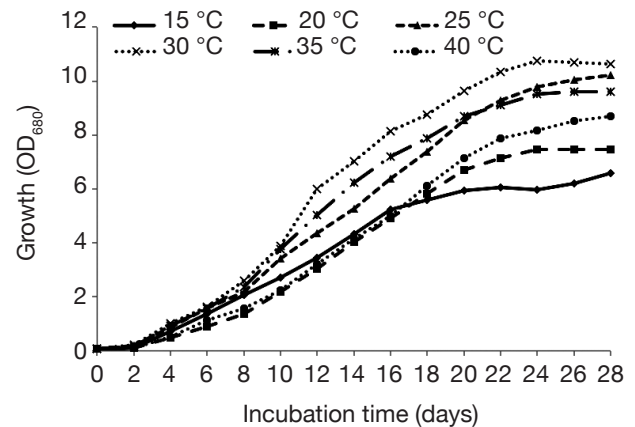
Results are presented as the mean  $\pm$  standard deviation (SD) of four replicates. Data were statistically analyzed using SAS (v 6.12). The degree of significance using one way analysis of variance and paired-samples t-test at probability level ( $p$ )  $\leq$  0.05 was determined.

## 3. RESULTS

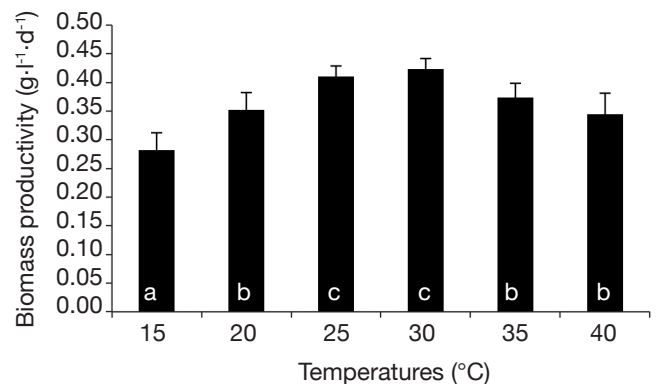
### 3.1. Growth and biomass productivity

Effect of incubation temperature on the growth rate of *S. acutus* was examined based on cell density (measured

as optical density) and cellular dry weight (biomass productivity) for 28 days of incubation (**Figure 1**). At late exponential phase (after 22 days), incubation at 30 °C showed the maximum growth, while the lowest growth was recorded at 15 °C. The growth reduction was less pronounced at 40 °C. Biomass productivity showed the same pattern, with significantly less (one way ANOVA,  $p \leq 0.05$ ) biomass productivity ( $0.28 \text{ g} \cdot \text{l}^{-1} \cdot \text{d}^{-1}$ ) at 15 °C (**Figure 2**), while the highest biomass productivity ( $0.42 \text{ g} \cdot \text{l}^{-1} \cdot \text{d}^{-1}$ ) was recorded at 30 °C. Differences in biomass productivity did not



**Figure 1.** Growth pattern of *Scenedesmus acutus* cultivated for 28 days at different temperatures — *Profils de croissance de Scenedesmus acutus suivis durant 28 jours à différentes températures.*



**Figure 2.** Effect of different growth temperatures on biomass productivity of *Scenedesmus acutus* — *Productivité en biomasse de Scenedesmus acutus à différentes températures de croissance.*

Error bars show the SD for four replicates — *la barre d'erreur représente l'écart-type sur quatre réplicats*; same letters indicate reported values without statistically significant difference ( $p \leq 0.05$ ) — *des lettres identiques associées aux valeurs rapportées indiquent que statistiquement leurs différences ne sont pas significatives* ( $p \leq 0,05$ ).

vary significantly between incubation temperatures of 25 and 30 °C. Incubation at 40 °C showed significant reduction in biomass productivity by 18% as compared with that at 30 °C (Figure 2).

### 3.2. Fatty acids content

The fatty acids contents per microalgal biomass ( $\text{mg}\cdot\text{g}^{-1}$  CDW) of *S. acutus* after 22 days of cultivation at different incubation temperatures are shown in figure 3. At 30 °C, *i.e.* at maximum growth rate, the fatty acids content was  $94.90 \text{ mg}\cdot\text{g}^{-1}$  CDW. At lower temperature (15 °C), the fatty acids content was  $104.12 \text{ mg}\cdot\text{g}^{-1}$  CDW, significantly (paired-samples t-test,  $p \leq 0.05$ ) higher than the one at 30 °C by 10%. At higher temperature (40 °C), the fatty acids content significantly decreased to  $86.34 \text{ mg}\cdot\text{g}^{-1}$  CDW.

### 3.3. Fatty acids productivity

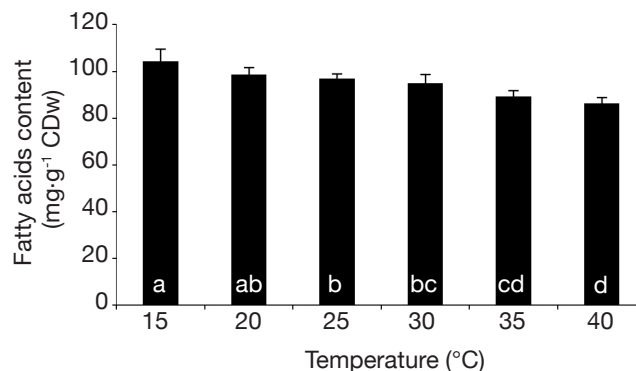
Esterified fatty acids (EFA) productivity of *S. acutus* at different cultivation temperatures is shown in figure 4. Due to the high microalgal biomass, the highest EFA productivity was recorded at 30 °C ( $42.10 \text{ mg}\cdot\text{l}^{-1}\cdot\text{d}^{-1}$ ). The lowest EFA productivity ( $\sim 30 \text{ mg}\cdot\text{l}^{-1}\cdot\text{d}^{-1}$ ) was achieved at 15 and 40 °C. The EFA productivity ( $\sim 35 \text{ mg}\cdot\text{l}^{-1}\cdot\text{d}^{-1}$ ) by *S. acutus* cultivated at 20 and 35 °C decreased by 29 and 17%, respectively, compared to that at 30 °C. Therefore, the optimal cultivation temperature to produce microalgal biomass (*S. acutus*) and fatty acids for biodiesel production ranges from 25 to 30 °C.

### 3.4. Fatty acids profile

Fatty acids composition of *S. acutus* grown at different temperatures is shown in table 1. The ratio of the sum of saturated and monounsaturated fatty acids increased by increasing the temperature, while under the same conditions, the ratio of polyunsaturated fatty acids was reduced. At relatively low temperature (15 °C), approximately 35% of total fatty acids of *S. acutus* was composed of polyunsaturated fatty acid (C18:3n-3). While at high temperature (40 °C), the fatty acids were mainly composed of the monounsaturated fatty acids (C18:1n-9c) and saturated fatty acid (C16:0) with 28 and 21%, respectively.

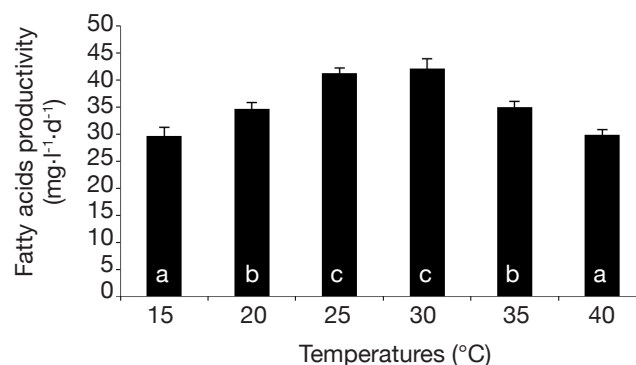
## 4. DISCUSSION

Microalgae are one of the potential sources for biodiesel production due to high efficiency of solar energy conversion to chemical energy. The impact of nutrient depletion and physical stress was examined on different algae for enhancement of lipid production. However, Abomohra et al. (2013) and Francisco et al.



**Figure 3.** Fatty acids content of *Scenedesmus acutus* after 22 days of incubation at different temperatures — *Teneur en acides gras de Scenedesmus acutus après 22 jours d'incubation à des températures différentes.*

Error bars show the SD for four replicates — *la barre d'erreur représente l'écart-type sur quatre réplicats*; same letters indicate that there is no statistically significant difference ( $p \leq 0.05$ ) — *des lettres identiques associées aux valeurs rapportées indiquent que statistiquement leurs différences ne sont pas significatives ( $p \leq 0,05$ ).*



**Figure 4.** Fatty acids productivity of *Scenedesmus acutus* at different temperatures — *Productivité en acides gras de Scenedesmus acutus à différentes températures de croissance.*

Error bars show the SD for four replicates — *la barre d'erreur représente l'écart-type sur quatre réplicats*; same letters indicate that there is no statistically significant difference ( $p \leq 0.05$ ) — *des lettres identiques associées aux valeurs rapportées indiquent que statistiquement leurs différences ne sont pas significatives ( $p \leq 0,05$ ).*

(2010) reported that biomass productivity and lipid content are inversely related. Therefore, for biodiesel production, the economic feasibility of microalgal mass culture has to be taken into consideration. The selection of microalgal species with high lipid contents and high cell growth is of great importance.

**Table 1.** Fatty acid profiles measured after 22 days of *Scenedesmus acutus* incubation at different temperatures — *Composition en acides gras de Scenedesmus acutus croissant à différentes températures, mesurée après 22 jours d'incubation.*

Fatty acid	Percent of fatty acids (% of total fatty acids)					
	15 °C	20 °C	25 °C	30 °C	35 °C	40 °C
C14:0	nd	0.20 ± 0.02	0.23 ± 0.01	0.27 ± 0.01	0.28 ± 0.01	0.26 ± 0.02
C16:0	13.31 ± 0.64	17.44 ± 1.80	19.88 ± 1.36	21.03 ± 1.12	19.74 ± 0.20	20.61 ± 1.32
C16:1n-10	1.14 ± 0.02	2.03 ± 0.13	1.72 ± 0.02	1.31 ± 0.09	1.36 ± 0.09	1.26 ± 0.14
C16:1n-7	3.20 ± 0.08	3.15 ± 0.15	2.76 ± 0.25	2.85 ± 0.56	3.34 ± 0.46	3.30 ± 0.55
C16:2	2.04 ± 0.03	1.82 ± 0.14	1.66 ± 0.20	1.63 ± 0.06	1.36 ± 0.09	1.34 ± 0.13
C16:3	8.31 ± 0.06	8.50 ± 0.59	6.62 ± 0.29	5.79 ± 0.72	4.94 ± 0.16	4.82 ± 0.23
C16:4n-3	14.71 ± 0.89	11.14 ± 0.75	7.96 ± 0.65	6.04 ± 0.29	5.05 ± 0.27	5.84 ± 0.32
C18:0	0.41 ± 0.12	0.88 ± 0.01	1.69 ± 0.10	1.91 ± 0.07	1.99 ± 0.06	2.27 ± 0.29
C18:1n-9t	nd	0.13 ± 0.01	0.18 ± 0.02	0.17 ± 0.02	0.17 ± 0.02	0.16 ± 0.01
C18:1n-9c	6.79 ± 0.10	15.07 ± 0.36	20.65 ± 0.65	26.16 ± 1.14	27.32 ± 1.50	28.38 ± 3.07
C18:02n-6	11.56 ± 0.13	11.89 ± 1.32	11.53 ± 0.76	11.67 ± 0.56	12.19 ± 0.37	10.84 ± 0.51
C18:3n-3	34.99 ± 2.16	23.13 ± 3.93	20.92 ± 1.49	17.04 ± 0.91	17.79 ± 0.38	16.49 ± 0.94
C18:3n-6	0.85 ± 0.08	1.54 ± 0.09	1.47 ± 0.01	1.22 ± 0.02	1.27 ± 0.03	1.18 ± 0.07
C18:4n-3	2.69 ± 0.09	3.07 ± 0.10	2.71 ± 0.09	2.69 ± 0.31	2.81 ± 0.33	2.61 ± 0.39
C20:0	nd	nd	nd	0.24 ± 0.01	0.38 ± 0.03	0.65 ± 0.03
SFA	13.72 ± 0.12 <sup>a</sup>	18.53 ± 0.82 <sup>b</sup>	21.81 ± 0.48 <sup>c</sup>	23.44 ± 1.19 <sup>d</sup>	22.39 ± 0.15 <sup>cd</sup>	23.79 ± 1.64 <sup>d</sup>
MUFA	11.13 ± 0.20 <sup>a</sup>	20.38 ± 0.63 <sup>b</sup>	25.31 ± 0.91 <sup>c</sup>	30.49 ± 0.69 <sup>d</sup>	32.20 ± 0.57 <sup>de</sup>	33.10 ± 2.60 <sup>e</sup>
PUFA	75.15 ± 0.14 <sup>a</sup>	61.09 ± 1.43 <sup>b</sup>	52.88 ± 1.38 <sup>c</sup>	46.07 ± 1.68 <sup>d</sup>	45.41 ± 0.62 <sup>de</sup>	43.10 ± 2.18 <sup>e</sup>

nd: not detected — *non détecté*; SFA: saturated fatty acid — *acide gras saturé*; MUFA: mono-unsaturated fatty acid — *acide gras mono-insaturé*; PUFA: poly-unsaturated fatty acids — *acide gras poly-insaturé*; values are given as percent (%) of total fatty acids — *les valeurs sont données en pourcentage des acides gras totaux*; values are the mean ± SD of four replicates — *les valeurs sont la moyenne de quatre répétitions ± écart-type*; values with the same letter in the same row indicate that there is no statistically significant difference ( $p \leq 0.05$ ) — *les valeurs avec la même lettre dans la même ligne indiquent qu'il n'y a pas de différence statistiquement significative ( $p \leq 0,05$ )*.

In the present study, the cell growth and consequently biomass productivity of *S. acutus* were closely related with cultivation temperature. The highest biomass productivity was achieved at 25–30 °C, while biomass productivity decreased constantly when increasing the temperature over 30 °C. Westerhoff et al. (2010) reported 20 °C as the optimal cultivation temperature for *Nannochloropsis oculata*. However, they concluded that the exponential growth rate constant did not vary between 27 and 39 °C for *Scenedesmus* and *Chlorella*, while at 42 °C algal growth stops.

Temperature is a sensitive parameter for microalgal growth and the concomitant metabolic activities, especially lipid biosynthesis. In the coupled system of wastewater treatment and biodiesel production by microalgae, increasing lipid productivity by temperature adjustment has the advantage that the composition of wastewater does not need to be changed or chemically treated for processing and is therefore ecologically safer (Xin et al., 2011). Chen et al. (2008) reported a

little effect of cultivation temperature on lipid content of microalgal biomass, with a significant decrease of triglycerides (TAGs) at low temperatures. However, Converti et al. (2009) reported that high temperature enhances the lipid accumulation in *Nostoc oculata*.

In our study, the dependence of lipid content of *S. acutus* on cultivation temperature is in agreement with the findings of Chen et al. (2008). In the present study, low temperature induced the fatty acid accumulation in *S. acutus* cells and resulted in high lipid content. From an economic viewpoint, enhancement of lipid productivity, which is related to the growth, has more feasibility than lipid content. Li et al. (2010) showed that high lipid contents of *Scenedesmus* sp. could be obtained under stress conditions; however due to low growth rate, the lipid productivity decreased as well as microalgal biomass productivity.

Abomohra et al. (2014) and El-Sheekh et al. (2017) studied the efficiency of pilot cultivation of *S. obliquus* as a feedstock for biodiesel. They recorded that the fatty

acids mass fraction in CDW and biomass production of *S. obliquus* after 21 days of pilot cultivation were 123.3 g·kg<sup>-1</sup> and 3.11 g·l<sup>-1</sup>, respectively. They concluded that the measured fatty acid productivity of 17.4 mg·l<sup>-1</sup>·d<sup>-1</sup> would be nearly 5 and 8 times higher than lipid productivity of *Jatropha* and rapeseed, respectively. However, fatty acid productivity ( $\approx$  42 mg·l<sup>-1</sup>·d<sup>-1</sup>) at the optimum temperature in the present study is about 50% greater than that recorded for *S. obliquus* by Abomohra et al. (2014). This confirms that microalgae might be a major biofuel feedstock with the potential to completely displace fossil diesel.

The saturation and chain length of fatty acids would affect the properties of biodiesel. Hu et al. (2008) mentioned that saturated fats produce a biodiesel with higher oxidative stability and cetane number, but rather poor low-temperature properties. However, biodiesel feedstock rich in polyunsaturated fatty acids produces a biodiesel with good cold-flow properties, but with high oxidation ability. In our study, lipids of *S. acutus* were mainly composed of polyunsaturated fatty acids at a relatively low cultivation temperature, but they mainly contained saturated and monounsaturated fatty acids at high temperatures. The fatty acid profile of *S. acutus* evolves with incubation temperature. It makes the biodiesel from microalgae suitable for cold or warm areas depending on the cultivation temperature under which the feedstocks are obtained.

## 5. CONCLUSIONS

In conclusion, the variation of temperature showed minor effects on lipid content of *S. acutus* and strongly affected the lipid productivity due to its influence on algal biomass production. The optimum temperature range for maximum lipid and fatty acids production was 25-30 °C. The degree of saturation of fatty acids increased with incubation temperature, which allows biodiesel properties from *S. acutus* to be used in different climates. Research on microalgae-based biofuel and commercial-scale use of microalgae for biofuel production would require massive investments because it is a highly promising way to meet the energy demand through third generation biofuel from the microalgal feedstock.

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